

Note on the intestinal bacterial populations of free living snakes in Italy

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As part of a survey of the herpetofauna in the "Riserva Naturale Regionale Selva del Lamone" protected area (Lazio Region, Central Italy), a bacteriological pilot study was carried out between March 23rd and June 10th 2008. Thereby, eleven snake specimens were captured and screened for cloacal pathogens.

The study area is located in Northern Lazio, at the border with Tuscany, and is characterized by a rich and diverse herpetofauna (Filippi et al., 2008). Distributed over a plain area (350 m a.s.l.) the dominant vegetation type consists of Turkey oak (*Quercus cerris*) with other deciduous trees, shrubs, arable land and pasture. The study area was surveyed for snakes along standardised routes, covering various micro-habitats. Snakes were captured by hand, and several parameters were recorded: a) date, site (GPS coordinates) and micro-habitat; b)

species; c) sex; d) snout-vent length and tail length.

Out of 77 captured snakes (see Table 1) eleven specimens belonging to three snake species were sampled by cloacal swab for bacteriological analysis: 7 Asp Vipers (*Vipera aspis*), 3 Western whip snakes (*Hierophis viridiflavus*), 1 Aesculapian snake (*Zamenis longissimus*).

Briefly, swabs were inoculated onto Brain Heart Infusion broth and incubated at 30 +/- 1 C° and 37 +/- 1 C° for 24 hours. Broths were then transferred onto blood and agar incubated as above. Colonies were identified by Gram's staining oxidase test, by inoculation onto Triple Sugar Iron and by API 20E system (Bio-Meurieux). Colonies identified as *Salmonella* spp. were then typified by the autoagglutination saline test, the polyvalent agglutination antisera test (A-O67) and by serological testing following the Kauffmann and White scheme (Carter and Cole, 1990; Holt et al., 1994; OIE, 2008).

The results of the bacteriological examination of the cloacal swabs and other data are reported in Table 2. All sampled snakes tested positive for a mixed Gram-negative bacterial flora. However, three *V. aspis* and one *Hierophis viridiflavus* yielded positive results for *Salmonella enterica diarizonae* IIIb.

Many reptiles harbor a mixed Gram-negative bacterial flora in their cloaca, and this presence is difficult to interpret. Bacteria, such as *Pseudomonas* spp., *Klebsiella* spp., *Serratia* spp. and *Providencia* spp., all of which have been isolated in this study, are all

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Table 1. Sampled snake population divided by species.

Species	<i>Hierophis viridiflavus</i>	<i>Zamenis longissimus</i>	<i>Elaphe quatuorlineata</i>	<i>Natrix natrix</i>	<i>Natrix tessellata</i>	<i>Vipera aspis</i>
Number of specimens	40	6	2	4	2	23

Table 2. Date, species, sex and size (Svl: snout-vent length (cm); Tl: tail length (cm)) and results of bacteriological analysis of cloacal swab

Date	Species	Sex	Svl	Tl	Isolates
23 March	<i>V. aspis</i>	M	50	62	<i>Hafnia alvei</i> ; <i>Salmonella enterica</i> sub. <i>diarizonae</i>
30 April	<i>H. viridiflavus</i>	F	90	122	<i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.; <i>Morganella morganii</i> ; <i>Buttiaux agrestis</i>
30 April	<i>H. viridiflavus</i>	F	71	99	<i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.; <i>Morganella morganii</i> ; <i>Buttiaux agrestis</i> ; <i>Lactococcus lactis</i>
30 April	<i>V. aspis</i>	F	40	46	<i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.; <i>Morganella morganii</i> ; <i>Buttiaux agrestis</i>
30 April	<i>V. aspis</i>	M	54	63	<i>Pseudomonas fluorescens</i> ; <i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.; <i>Morganella morganii</i> ; <i>Buttiaux agrestis</i>
07 May	<i>V. aspis</i>	M	43	47.5	<i>Klebsiella pneumoniae</i> ; <i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.
07 May	<i>V. aspis</i>	F	57	--	<i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.
07 May	<i>Z. longissimus</i>	M	94	117	<i>Serratia liquefaciens</i> ; <i>Citrobacter</i> spp.
10 June	<i>V. aspis</i>	F	46	51.5	<i>Escherichia coli</i> ; <i>Corynebacterium</i> sp.; <i>Providencia rettgeri</i> ; <i>Salmonella enterica</i> sub. <i>diarizonae</i>
10 June	<i>V. aspis</i>	M	49	56	<i>Salmonella enterica</i> sub. <i>diarizonae</i> ; <i>Providencia rettgeri</i> ;
10 June	<i>H. viridiflavus</i>	F	84	111	<i>Salmonella enterica</i> sub. <i>diarizonae</i> ; <i>Providencia rettgeri</i> ; <i>Corynebacterium</i> spp.; <i>Pasteurella pneumotropica</i>

considered opportunistic pathogens in reptiles, and their presence in association with overt clinical signs should be considered as significant (Rosenthal and Mader, 1996).

Salmonella enterica has been previously isolated from free-living snakes (Malmqvist *et al.*, 1995; Geue and Löschner, 2002; Briones *et al.*, 2004; Köbölkuti *et al.*, 2009), captive snakes (Kennedy, 1973; Chiodini and Sundberg, 1981; Geue and Löschner, 2002; Schroter *et al.*, 2004) and their environment in captivity (Bauwens *et al.*, 2006), including pet snakes in Italy (Ebani *et al.*, 2005).

However, to the best of the authors' knowledge these are the first reported *Salmonella* isolates from wild specimens of *Vipera aspis* and *Hierophis viridiflavus*.

Salmonella species are potential pathogens for reptiles in general (Funk, 1996), and *Salmonella enterica diarizonae* is a rarely reported zoonotic agent (Bruins *et al.*, 2006).

The present study, which was carried out by conservation biologists and veterinary health professionals, highlighted a potential biological threat to the snakes under survey. Additionally, the role of wild snakes as a reservoir of *Salmonella* species was

confirmed, hence the need to apply strict hygienic precautions when handling them.

Considering the decline of many snake populations (Reading et al., 2010) and that the spreading of diseases is considered to be one of the causes for the decline in reptiles in general (Gibbons et al., 2000), it is important to remember that “today’s commensal may be tomorrow’s pathogen” (Cooper, 2000).

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