

Reproductive diversity of Malawian anurans

Vincenzo Mercurio^{1,2*}, Wolfgang Böhme³, Bruno Streit¹

Abstract. Of the 39 described reproductive modes of anurans, few are recognisable among the anuran species of Malawi. The reproductive modes recognised in Malawi anurans are: i) the deposition of aquatic eggs and free feeding tadpoles in lentic water (Mode 1 sensu Kentwood, 2007), e.g. as present in the genera *Amietophrynus*, *Phrynobatrachus*, *Ptychadena*, and *Xenopus* comprising 72% of the species; ii) the deposition of eggs in leaves over water hatching into free feeding tadpoles that drop in lentic water (Mode 24), genera *Afrixalus* and *Hyperolius* representing 13% of the species; iii) direct development of terrestrial eggs (Mode 23) in the genera *Arthroleptis* and *Breviceps*, 8% of the species; iv) terrestrial eggs deposited on ground above water hatching in free feeding tadpoles which move to water (Mode 18) genera *Leptopelis* and *Hemisus*, 4% of the species; v) eggs deposited above water on humid rocks or in rock crevices, hatching into free feeding semiterrestrial tadpoles living on rocks in a water film (Mode 19), two species in two genera *Ptychadena broadleyi* and *Nothophryne broadleyi*, 2% of the species; vi) arboreal foam nest hatching into free feeding tadpoles that drop into ponds or streams (Mode 33), only the species *Chiromantis xerampelina*, 1%. The main characteristic of the Malawian anuran fauna is the low richness of reproductive modes, the predominance of habitat generalist species, and the high similarities of habitats across the country. Among the six occurring reproductive modes, five (Mode 1, 18, 24, 23, and 33) are widespread in Malawi and commonly found in most communities, whereas Mode 19 is restricted to two species occurring at Mulanje and Zomba mountains.

Keywords. amphibians, diversity, Malawi, model animals, reproductive modes.

There have been several attempts to classify the reproductive modes of amphibians (Salthe and Duellman, 1973; Duellmann and Trueb, 1986; Haddad and Prado, 2005; Kentwood, 2007). The definition of different reproduction modes embraces the combination of several reproductive and developmental characters including site of oviposition, shape and size of the egg clutch, kind and duration of development, tadpole type, size at metamorphosis, and parental care (Duellmann and Trueb, 1986; Haddad and Prado, 2005; Kentwood, 2007).

Among vertebrates, amphibians show the highest level of diversification of reproductive strategies, and at the same time they are extremely dependent by the availability of water (Duellmann and Trueb, 1986;

McDiarmid, 1994; Vences and Köhler, 2008). Many of these reproductive strategies can be interpreted as an evolutionary trend towards the progressive emancipation from water, allowing reproduction in more terrestrial habitats (Bogart, 1981; Duellman, 1985; Duellman and Trueb, 1986; Lutz, 1947; Salthe and Mecham, 1974). However, the selective pressure (e.g. protection against predators, reduction of larval competition, escape from desiccation, etc) leading to this evolutionary trend is still object of debate (Kentwood, 2007; Martin, 1967; Van Dijk, 1971).

Altogether a total of 39 reproductive modes were described for amphibians with most of them to be found in tropical areas, especially in the New World, and only 12 reproductive modes were listed for sub-Saharan Africa (Duellmann and Trueb, 1986; Haddad and Prado, 2005; Kentwood, 2007). In general, high degrees of diversity of reproductive modes is found in lowland rain forests with high niche heterogeneity, whereas it typically decreases in arid homogeneous regions where generalist pond breeders usually prevail (dos Santos et al. 2008, 2009; Duellmann and Trueb, 1986; Keller et al. 2009; Kentwood, 2007; Perotti, 1997; Prado et al. 2005). Consequently, highly diverse communities are much more sensible to habitat alteration than generalised ones,

1 Department of Ecology and Evolution, Goethe-Universität, Biologie-Campus, Siesmayerstr. 70, 60323 Frankfurt am Main, Germany;

2 Forschungsinstitut und Naturhistorisches Museum Senckenberg, Sektion Herpetologie, Senckenberganlage 25, 60325, Frankfurt am Main, Germany;
e-mail: vincenzomercurio@gmx.de

3 Zoologisches Forschungsmuseum Alexander Koenig, Adenauerallee 160, D-53113 Bonn, Germany.

* corresponding author

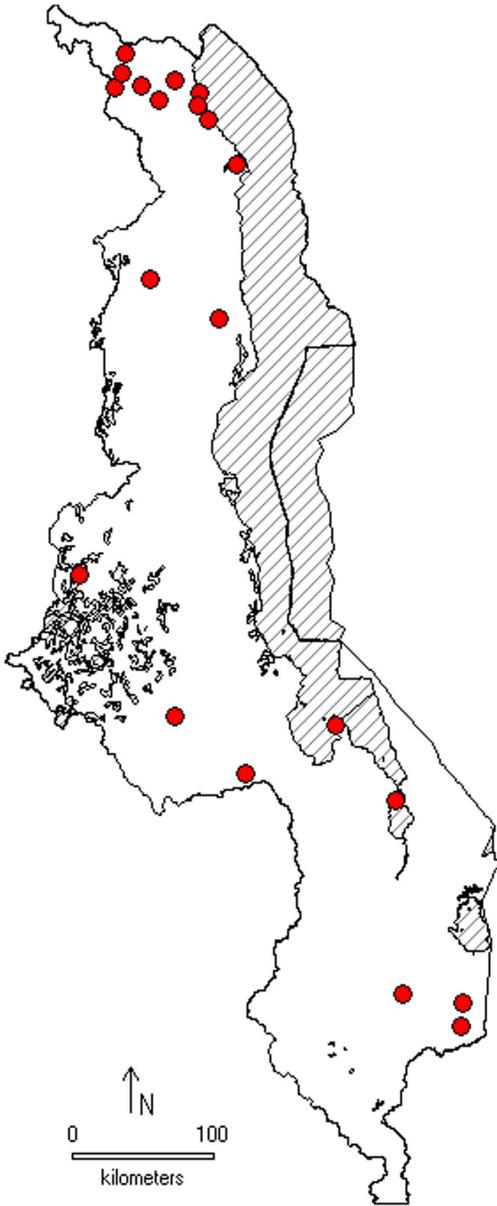


Figure 1. Schematic map of Malawi showing the distribution of the main studied sites.

and the degree of diversification of reproductive modes can be used to roughly predict the possible reactions of communities to habitat modification (Haddad and Prado, 2005; Mercurio, 2009).

Detailed ecological data about African amphibians are scarce (Amiet, 1972; Channing, 2001; Channing and Howell, 2006; Passmore and Carruthers, 1995; Rödel, 1996, 2000; Stewart, 1967; Schiøtz, 1999; Van Dijk,

1972, 1977; Wager, 1965). In many cases available information about species are anecdotal and based on few, or even single, isolated observations. Descriptions about egg deposition sites, clutch size and shape, duration of development, and tadpoles lacks for many African species, as is the case for several Malawian anurans. Any attempt of a comprehensive ecological treatment will be hampered by this absence of data. Therefore, for Malawi, whenever reasonable, baseline ecological information (e.g. eggs deposition site; presence of exotrophic tadpole) have been inferred from closely related species.

Field trips were carried out in four periods: (1) 27th January–9th March 2006; (2) 1st February–30th April 2007; (3) 3rd November–10th December 2007; (4) 19th January–17th March 2008, with a total of 115 days of field activity and 25 visited sites (Fig. 1). Daily search consisted of searching at all hiding places and suitable microhabitats available at the study site (under rocks, inside logs, within leaf litter, under loose bark, and dead tree branches). During the night frogs were located with the aid of flashlights and headlamps, either by opportunistic searches or by the localisation of calling males. Permanence in the study area varied between one and 15 days.

Following the classification proposed by Kentwood (2007), who modified the scheme of Haddad and Prado (2005), six reproductive modes are recognisable in the anuran species of Malawi (Fig. 2); listed from the most common to the rarest with the relative percentage of the fauna: i) aquatic eggs and free feeding tadpoles in lentic water (Mode 1), e.g. genera *Amietophrynus*, *Phrynobatrachus*, *Ptychadena*, and *Xenopus*, 72%; ii) eggs in leaves over water hatching into free feeding tadpoles that drop in lentic water (Mode 24), genera *Afrixalus* and *Hyperolius*, 13%; iii) direct development of terrestrial eggs (Mode 23), genera *Arthroleptis* and *Breviceps*, 8%; iv) terrestrial eggs on ground above water hatching in free feeding tadpoles which move to water (Mode 18), genera *Leptopelis* and *Hemisus*, 4%; v) eggs above water on humid rocks or in rock crevices, hatching into free feeding semiterrestrial tadpoles living on rocks in a water film (Mode 19), two species in two genera *Ptychadena broadleyi* and *Nothophryne broadleyi*, 2%; vi) arboreal foam nest hatching into free feeding tadpoles that drop into ponds or streams (Mode 33), only the species *Chiromantis xerampelina*, 1%. The known reproductive data of Malawian anurans and their allocation to a reproductive mode sensu Kentwood (2007) is summarised in Table 1.

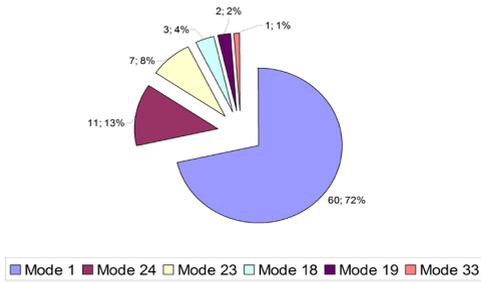


Figure 2. Reproductive modes in Malawian anurans. Occurrence, relative number of species, and percentage of the six reproductive modes. Most of the species reproduce by means of Mode 1.

Mode 1: Pond breeders with free feeding tadpole. This reproductive mode is the most generalised and phylogenetically widespread one, being found worldwide in most of the anuran families (Duellmann and Trueb, 1986; Kentwood, 2007). It is characterised by the deposition of eggs in temporary water bodies which develop into exotrophic aquatic tadpole until metamorphosis (Haddad and Prado, 2005). Of the 12 anuran families which occur in Malawi eight families (66%) and 17 genera (74%) share this reproductive mode. Of the 84 anuran species about two thirds (60; 72%) are generalist pond breeders, and most of them have a benthic tadpole (Table 1). However, despite the majority of Malawian anurans are pond breeders and their reproduction can be adequately described by the Mode 1, the species show some relevant differences in their breeding behaviour including: i) the nature and size of the oviposition sites, ii) clutch size iii) and at lesser extent tadpole type. Nearly all the species within this category normally breed in temporary lentic water bodies of various sizes, and only few species occasionally breed in lotic waters. Examples are the species belonging to the genera *Amietia*, *Amietophrynus*, *Hylarana*, *Ptychadena*, and *Strongylopus* which also breed in stagnant pools within streams with slow moving water. Further differences are evident in the size of the breeding site. Due to the paucity of data and the intrinsic ecological plasticity of Mode 1 it is difficult to categorise these species on the basis of their preferences about breeding site sizes. One exception is represented by the genus *Phrynobatrachus* which often breeds in very small rain puddles, ditches, and road drainages with shallow water. Species with Mode 1 lay their eggs directly in water. In Malawi, one exception is represented by *Hyperolius argus* for which the eggs were reported to be deposited under water, attached to submerged vegetation (Wagner,

1985). However, it is not sure whether this represents a real trait of the reproductive mode or rather the eggs were submerged due to the raising of the water level after heavy rains (Schlotz, 1999). Relevant differences among Mode 1 species are present in the clutch sizes. Some species like *Ptychadena taenioscelis* and *Strongylopus merumontanus* lay very small clutches with 2–10 and 1–14 eggs respectively, whereas species like *Amietophrynus garmani* and *A. gutturalis* are able to produce clutches containing up to 25000 eggs (Channing, 2001; Stewart, 1967). Unfortunately for 36 species (60%) detail about the egg clutch size are unknown. Channing (2001) reported the clutch size of *Mertensophryne taitana* being composed of 125 eggs with the citation of Stewart (1967). However, no direct information is available for this species in the above mentioned work and 125 eggs were instead reported for *Mertensophryne nyikae*. A total number of 54 species have free feeding benthic tadpoles comprising 90% of this category. Exceptions are two species belonging to the genus *Kassina* (*K. maculata*, and *K. senegalensis*) and the toad *Schismaderma carens*, both with nektonic tadpoles; two *Xenopus* species (*X. laevis* and *X. muelleri*) and *Phrynomantis bifasciatus* both with suspension feeder tadpoles. Within this category one species *Pyxicephalus adspersus* shows some parental care behaviour. Males of this species dig channels for the tadpoles trapped in drying marginal puddles preventing their death through desiccations (Kok, Du Preez and Channing, 1989; Channing, Du Preez and Passmore, 1994). Of the 60 species reproducing by means of Mode 1, 48 species (80%) were found breeding in natural as well as secondary habitats, farm bush landscape and gardens, three species (5%) *Amietophrynus kisoensis*, *Mertensophryne nyikae*, and *Strongylopus merumontanus* are apparently linked with the presence of a specific habitat, and the remaining nine species (15%) are unclassifiable due to lack of data. In summary, most of the Malawian amphibians are generalised pond breeders showing a considerable ecological plasticity being able to reproduce in significantly different habitat conditions.

Mode 24: Eggs deposited in leaves over water with free feeding tadpole. In Malawi 11 species (13%) belonging to the genera *Afraxalus* and *Hyperolius* (family Hyperoliidae) are to be found with this reproductive mode (Table 1). These species breed in temporary lentic body waters of various sizes in open habitat, and deposit the eggs in the vegetation above water. However, in absence of vegetation the direct oviposition in water occasionally

Table 1. Reproductive and developmental characters of the anurans of Malawi. Ecological classification of tadpoles follows McDiarmid and Altig (1999). Clutch size and development data are taken from Channing (2001), Channing and Howell (2006), Rödel (2000), Schiötz (1999), Stevens (1972), Stewart (1967), and Wager, (1965). The terms “generalist” describes a species known to breed in natural, as well as in perturbed, whereas the term “specialist” describes species known to breed in natural and unperturbed areas only. RM = classification of reproductive modes after Haddad and Prado (2005) and Kentwood (2007). See text for further details.

Species	Eggs Deposition	Water	Clutch size	Development	Tadpole	Category	RM
Arthroleptidae							
<i>Arthroleptis francei</i>	leaf litter	—	unknown	direct	—	unknown	23
<i>Arthroleptis reichei</i>	leaf litter	—	unknown	direct	—	unknown	23
<i>Arthroleptis stenodactylus</i>	leaf litter	—	33–80	direct	—	generalist	23
<i>Arthroleptis xenochirus</i>	leaf litter	—	9	direct	—	unknown	23
<i>Arthroleptis xenodactyloides</i>	leaf litter	—	20	direct	—	generalist	23
<i>Leptopelis argenteus</i>	water	lentic	unknown	tadpole	benthic	unknown	1
<i>Leptopelis bocagii</i>	hole underground	lentic	unknown	tadpole	benthic	unknown	18
<i>Leptopelis flavomaculatus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Leptopelis mossambicus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Leptopelis parbocagii</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Leptopelis sp.</i>	water	lentic	unknown	tadpole	benthic	unknown	1
Brevicipitidae							
<i>Breviceps mossambicus</i>	subterranean nest	—	20–30	direct	—	generalist	23
<i>Breviceps poweri</i>	subterranean nest	—	unknown	direct	—	unknown	23
Bufonidae							
<i>Amietophrynus garmani</i>	water	lentic	12000–20000	tadpole	benthic	generalist	1
<i>Amietophrynus gutturalis</i>	water	lentic, lotic	15000–25000	tadpole	benthic	generalist	1
<i>Amietophrynus kisoensis</i>	water	lentic, lotic	unknown	tadpole	benthic	specialist	1
<i>Amietophrynus maculatus</i>	water	lentic, lotic	2000–8000	tadpole	benthic	generalist	1
<i>Mertensophryne lindneri</i>	water	lentic, lotic	unknown	tadpole	benthic	generalist	1
<i>Mertensophryne nyikae</i>	water	lentic	125	tadpole	benthic	specialist	1
<i>Mertensophryne taitana</i>	water	lentic	125	tadpole	benthic	generalist	1
<i>Poyntonophrynus beiranus</i>	water	lentic	unknown	tadpole	benthic	unknown	1
<i>Schismaderma carens</i>	water	lentic	2500	tadpole	nektonic	generalist	1
Hemisotidae							
<i>Hemisis guineensis</i>	hole underground	lentic	unknown	tadpole	nektonic	generalist	18
<i>Hemisis marmoratus</i>	hole underground	lentic	80–250	tadpole	nektonic	generalist	18
Hyperolidae							
<i>Afrixalus brachycnemis</i>	vegetation above water	lentic	50	tadpole	benthic	generalist	24
<i>Afrixalus crotalus</i>	folded leaf above water	lentic	50	tadpole	benthic	generalist	24
<i>Afrixalus delicatus</i>	folded leaf above water	lentic	50	tadpole	benthic	generalist	24

Table 1. (continued)

<i>Afrixalus fornasini</i>	folded leaf above water	lentic	80	tadpole	benthic	generalist	24
<i>Hyperolius argus</i>	under water	lentic	200	tadpole	benthic	generalist	1
<i>Hyperolius kachalolae</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Hyperolius kivuensis</i>	vegetation above water	lentic	unknown	tadpole	benthic	generalist	24
<i>Hyperolius m. marginatus</i>	water	lentic	390	tadpole	benthic	generalist	1
<i>Hyperolius m. taeniatus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Hyperolius minutissimus</i>	water	lentic	unknown	tadpole	benthic	unknown	1
<i>Hyperolius mitchelli</i>	vegetation above water	lentic	50–100	tadpole	benthic	generalist	24
<i>Hyperolius nasutus</i> complex	water	lentic	200	tadpole	benthic	generalist	1
<i>Hyperolius pictus</i>	vegetation above water	lentic	60–90	tadpole	benthic	generalist	24
<i>Hyperolius pusillus</i>	water	lentic	20–120	tadpole	benthic	generalist	1
<i>Hyperolius quinquevittatus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Hyperolius spinigularis</i>	vegetation above water	lentic	150–200	tadpole	benthic	generalist	24
<i>Hyperolius substriatus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Hyperolius tuberilinguis</i>	vegetation above water	lentic	236–400	tadpole	benthic	generalist	24
<i>Hyperolius v. albofasciatus</i>	vegetation above water	lentic	515	tadpole	benthic	generalist	24
<i>Hyperolius v. nyassae</i>	vegetation above water	lentic	369	tadpole	benthic	generalist	24
<i>Kassina maculata</i>	water	lentic	unknown	tadpole	nektonic	generalist	1
<i>Kassina senegalensis</i>	water	lentic	260–400	tadpole	nektonic	generalist	1
Microhylidae							
<i>Phrynomantis bifasciatus</i>	water	lentic	400–1500	tadpole	suspension feeder	generalist	1
Phrynobatrachidae							
<i>Phrynobatrachus acridoides</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Phrynobatrachus mababiensis</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Phrynobatrachus natalensis</i>	water	lentic	200–400	tadpole	benthic	generalist	1
<i>Phrynobatrachus parvulus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Phrynobatrachus perpalmatus</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Phrynobatrachus rungwensis</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Phrynobatrachus sp.</i>	water	lentic	unknown	tadpole	benthic	unknown	1
<i>Phrynobatrachus stewartae</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Phrynobatrachus ukingensis</i>	water	lentic	unknown	tadpole	benthic	generalist	1
Pipidae							
<i>Xenopus laevis</i>	water	lentic	1000	tadpole	suspension feeder	generalist	1
<i>Xenopus muelleri</i>	water	lentic	unknown	tadpole	suspension feeder	generalist	1

Table 1. (continued)

Ptychadenidae							
<i>Hildebrandtia ornata</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Ptychadena anchietae</i>	water	lentic	200–300	tadpole	benthic	generalist	1
<i>Ptychadena ansorgii</i>	water	lentic, lotic	unknown	tadpole	benthic	unknown	1
<i>Ptychadena broadleyi</i>	water	lentic	unknown	tadpole	semi- terrestrial	specialist	19
<i>Ptychadena guibei</i>	water	lentic	unknown	tadpole	benthic	unknown	1
<i>Ptychadena mascareniensis</i>	water	lentic	1080	tadpole	benthic	generalist	1
<i>Ptychadena mossambica</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Ptychadena oxyrhynchus</i>	water	lentic	3476	tadpole	benthic	generalist	1
<i>Ptychadena porosissima</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Ptychadena schillukorum</i>	water	lentic	unknown	tadpole	benthic	generalist	1
<i>Ptychadena taenioscelis</i>	water	lentic	2–10	tadpole	benthic	generalist	1
<i>Ptychadena upembae</i>	water	lentic	unknown	tadpole	benthic	unknown	1
<i>Ptychadena uzungwensis</i>	water	lentic	unknown	tadpole	benthic	unknown	1
Pyxicephalidae							
<i>Amietia angolensis</i>	water	lotic	unknown	tadpole	benthic	generalist	1
<i>Amietia cf. fuscigula</i>	water	lotic	unknown	tadpole	benthic	generalist	1
<i>Amietia johnstoni</i>	water	lotic, lentic	unknown	tadpole	benthic	generalist	1
<i>Amietia viridireticulata</i>	water	lotic, lentic	unknown	tadpole	benthic	unknown	1
<i>Nothophryne broadleyi</i>	wet moss	lentic	30	tadpole	unknown	specialist	19
<i>Pyxicephalus adspersus</i>	water	lentic	3000–4000	tadpole	benthic	generalist	1
<i>Strongylopus merumontanus</i>	water	lotic	1–14	tadpole	benthic	specialist	1
<i>Tomopterna cryptotis</i>	water	lentic	2000–3000	tadpole	benthic	generalist	1
<i>Tomopterna marmorata</i>	water	lentic	150	tadpole	benthic	generalist	1
<i>Tomopterna tuberculosa</i>	water	lentic	unknown	tadpole	benthic	generalist	1
Rhacophoridae							
<i>Chromantis xerampelina</i>	water	lentic	500–1250	tadpole	benthic	specialist	33
Ranidae							
<i>Hylarana darlingi</i>	water	lotic, lentic	unknown	tadpole	benthic	generalist	1
<i>Hylarana galamensis</i>	water	lentic	1500–4000	tadpole	benthic	generalist	1

occur (Pickersgill, 2007). *Afrivalus* species are used to lay the eggs above the water level in folded leaves glued together (Channing, 2001, Pickersgill, 1984, Schiötz, 1999). Apparently, one exception is represented by *A. brachynemis* where the eggs are not deposited in glued leaves (Channing, 2001). Clutch sizes are different

among those genera. *Afrivalus* species lay up to 50–80 eggs, whereas *Hyperolius* species lay up to about 500 eggs (Channing, 2001, Schiötz, 1999, Stewart, 1967; Pickersgill, 2007). They all share a generalised tadpole of the benthic type (McDiarmid and Altig, 1999). All the above mentioned Mode 24 species were found breeding

in natural as well as secondary habitats, e.g., farm bush landscape and gardens whenever suitable water bodies were available.

Mode 23: Terrestrial nests breeders with direct development. Of the 12 anuran families which occur in Malawi two families (17%) and two genera (8.7%) share this reproductive mode (Table 1). Altogether, seven species (8.2%) belonging to two genera *Arthroleptis* (*A. francei*, *A. reichei*, *A. stenodactylus*, *A. xenochirus*, and *A. xenodactyloides*), and *Breviceps* (*B. mossambicus*, and *B. poweri*) lay their eggs in holes underground or in the leaf litter, from which metamorphosed froglets hatch directly (Barbault and Rodrigues, 1979; Channing, 2001; Guibé and Lamotte, 1958; Lamotte and Perret, 1963; Thibaudeau and Altig, 1999; Wager, 1960, 1965). These species share a small clutch size with clutches containing up to 80 eggs in *A. stenodactylus* and up to 30 eggs in *B. mossambicus* (Channing, 2001; Stewart, 1967). The direct development allows these frogs to breed in a variety of very different habitats provided a certain amount of soil moisture is available. Three of these species (*A. stenodactylus*, *A. xenodactyloides*, and *Breviceps mossambicus*) can be defined as broad habitat generalists occurring in near all available habitats including cultivated areas and villages. Two of them are geographically restricted to small areas: *Arthroleptis francei* (Mulanje Mountain), and *A. reichei* (Misuku Mountains), but were collected in primary habitats (evergreen mountain forest, grassland, and shrubs) as well as in secondary habitats (e.g., tree plantations). Further studies are essential to assess the ecological plasticity of these species and their ability to survive in altered landscapes. The remaining two species *A. xenochirus* and *B. poweri* are poorly known.

Mode 18: Nest breeders with free feeding tadpole. Of the 12 anuran families occurring in Malawi two families (17%) and two genera (8.7%) share this reproductive mode (Table 1). Three species (3.5%) belonging to *Leptopelis* (*L. bocagii*) and *Hemisis* (*H. guineensis*, *H. marmoratus*) lay their eggs in holes underground close to water bodies where they develop until tadpoles hatch. Subsequently, in the *Leptopelis* species the tadpoles hatch and wriggle into the water, whereas the *Hemisis* tadpoles reach water through a channel dug by the female (Channing, 2001; Channing and Howell, 2006; Lamotte and Perret, 1961; Largen, 1977; Rödel, 1996, 2000; Rödel et al., 1995; Schiötz, 1999; Wager, 1965). Often they are simply flooded away by rain. Within this category clutch size is known only for *H. marmoratus* comprising 80–250 eggs (Channing, 2001; Rödel, 2000;

Wager, 1986). Concerning the tadpoles, *L. bocagii* have a benthic tadpole whereas the two *Hemisis* species have nektonic tadpoles (McDiarmid and Altig, 1999). These three species breed in temporary lentic body waters of various sizes. *H. marmoratus* is known to breed within farm bush landscape and gardens. The remaining two species *L. bocagii* and *H. guineensis* are poorly known.

Mode 19: Humid rocks breeders with semiterrestrial tadpoles. Only two species (2%) *Ptychadena broadleyi* (Ptychadenidae) and *Nothophryne broadleyi* (Pyxicephalidae) are found in Malawi with this peculiar reproductive mode (Table 1). *Ptychadena broadleyi* was reported as a completely rupicolous species with eggs deposited in pools within rocky outcrops from which semiterrestrial tadpoles hatch (Stevens, 1972). Details of oviposition site and clutch size are unknown. Stevens (1972) suggested the presence of aquatic eggs deposited as a floating mass in a single layer. Apparently, the tadpoles of *P. broadleyi* are unique within the genus being able to develop on sun exposed rock faces with only a film of water and the greater part of their bodies out of water (Stevens, 1972; Channing, 2001). They are adapted to move and adhering to the rock surface by means of the mouth and tail, feeding on algae and bacteria growing in the film of water (Channing, 2001; Stevens, 1972). Tadpole morphology differs from that of typical *Ptychadena* of the benthic type in having an elongate body and tail, with low fins resembling those of the genus *Leptopelis* (Stevens, 1972). Very little is known about the breeding behaviour of *Nothophryne broadleyi*. For this species Mazibuko and Poynton (2004) reported that “it is particularly associated with rocky areas in both montane forest and grassland. The eggs are laid in wet moss at the edge of rivulets running over rocks, and the larvae disperse by migration across wet rocks”. Both species are restricted to the plateaux of Mulanje and Zomba Mountains at 900–2715 m altitudes and giving their peculiar reproductive mode are here defined as habitat specialist. However, *P. broadleyi* was found in rocky outcrops within forest, clearings, as well as within gardens and tree plantations (Stevens, 1972). Due to the limited distribution both species are considered as endangered (IUCN, 2008). Further studies are essential to assess the status of these species and their ability to survive in altered landscapes.

Mode 33: Foam nest breeders in trees. Of the 12 anuran families occurring in Malawi only one species belonging to the Rhacophoridae built foam nests. *Chiromantis xerampelina* deposit the eggs in foam nest overhanging different kinds of lentic and lotic water

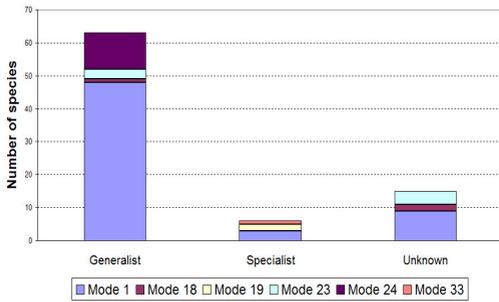


Figure 5. Ecological categorisation of the anuran of Malawi. The graphic shows the number of species in the six different reproductive modes. Habitat generalists are species known to thrive both in primary and altered habitat. Habitat specialists are known to breed only in primary habitat. Unknown, no data available. Most of the anurans of Malawi (75%) can be defined as habitat generalist reproducing in lentic temporary waters and able to survive in altered habitats. See text for further explanations.

bodies (Table 1). Subsequently, tadpoles drop in the water where they feed until metamorphosis is completed (Wager, 1926, 1965; Coe, 1974; Jennions, Backwell and Passmore, 1992; Schiötz, 1999). Usually *C. xerampelina* lays the eggs in foam nests hanging from branches of trees above water at different heights, but in absence of trees the species lay the eggs in any suitable objects. Therefore, this species shows a remarkable plasticity concerning the choice of oviposition sites. The tadpole is of benthic type (McDiarmid and Altig, 1999). Clutch size comprises 500-1250 eggs (Channing, 2001; Stewart, 1967). *Chiromantis xerampelina* is found in a variety of dry habitats from dry forest, miombo, mopane, savannah, to cultivated areas at low altitudes. However, this species appear to be much common in light wooded savannahs and forested areas. In fact, despite the occasional deposition of eggs directly in water or in grassy vegetation above water, the presence of a vegetation type able to support the large body size of the adults probably plays an important role in determining the occurrence of this species. Therefore, *C. xerampelina* is here preliminarily considered as a habitat specialist.

One main character of the Malawian anuran fauna is the low diversity of reproductive modes, the predominance of habitat generalist species, and the high similarity among used habitats. Among the six observed reproductive modes, five (Mode 1, 18, 24, 23, and 33) are widespread in Malawi and commonly found in most communities, whereas Mode 19 is restricted to two species occurring at Mulanje and Zomba mountains at medium to high altitudes. Furthermore, based on the

above mentioned classification and also taking into account the occurrence of many species in different primary and secondary habitats (Mercurio, 2009), 75% of the anuran fauna can be considered as habitat generalist (Fig. 3). As a consequence, these amphibians possess a scarce capacity to act as surrogates of habitat integrity and their use as model animals to detect habitat changes by means of quantitative studies should be valued case by case.

Acknowledgements. This work benefited from the financial support of the Deutsche Gesellschaft für Herpetologie und Terrarienkunde (DGHT), Hans-Schiemenz-Fonds, and from the Deutscher Akademischer Austausch Dienst (DAAD). A sincere and grateful thank you goes to Andrea Guerrini (Siena) for generously supporting me in a very critical moment of the research. I am indebted to the team of the Cultural and Museum Centre Karonga (CMCK), of the National Museum of Malawi (MM), the Department of Parks and Wildlife (DPW), and of the National Research Council of Malawi (NRCM) for their help in the field, logistic support, and for issuing the necessary research permits. A special thank goes to Jörn Köhler for his help in form of fruitful discussion and valuable comments.

References

- Amiet, J.-L. (1972): Quelques aspects de la biologie des amphibiens anoures du Cameroun. *Ann. Biol.* **28**: 73–136.
- Barbault, R., Rodrigues, M.T. (1979): Observations sur la reproduction et la dynamique des populations de quelques anoures tropicaux. 3. *Arthroleptis poecilnotus*. *Trop. Ecol.*, **20**(1): 64–77.
- Bogart, J.P. (1981): How many times has terrestrial breeding evolved in anuran amphibians? *Monit. Zool. Ital.*, n.s. suppl. **15**: 29–40.
- Channing, A.E. (2001): *Amphibians of Central and Southern Africa*. Comstock Publishing Associates, Ithaca and London, pp. 470.
- Channing, A.E., Du Preez, L., Passmore N. (1994): Status, vocalization and breeding biology of two species of African bullfrogs (Ranidae, *Pyxicephalus*). *J. Zool. (Lond.)*, **234**(1): 141–148.
- Channing, A.E., Howell, K.M. (2006): *Amphibians of East Africa*. Cornell University Press and Edition Chimaira, Ithaca.
- Coe, M.J. (1974): Observations on the ecology and breeding biology of the genus *Chiromantis* (Amphibia: Rhacophoridae). *J. Zool. (Lond.)*, **172**: 13–34.
- Dos Santos, T.G., Kopp, K., Spies, M.R., Trevisan, R., Cechin, S.Z. (2008): Temporal and spatial distribution of anurans in the Pampa Region (Santa Maria, RS). *Iheringia Sér. Zool.*, **98**(2): 244–253.
- Dos Santos, T.G., Vasconcelos, T.D., Rossa-Feres D.D., Hadad, C.F.B. (2009): *Anurans of a seasonally dry tropical forest: Morro do Diabo State Park, Sao Paulo state*. *J. Nat. Hist.*, **43**(15-16): 973–993.
- Duellman, W.E. (1985): Reproductive modes in anuran amphibians: Phylogenetic significance of adaptive strategies. *S. Afr. J. Sci.*, **81**: 174–178.

- Duellman, W.E., Trueb, L. (1986): *Biology of Amphibians*. McGraw-Hill, New York.
- Guibé, J., Lamotte, M. (1958): **Morphologie et reproduction par développement direct d'un anoure du Mont Nimba, *Arthroleptis crusculum* Angel.** Bull. Mus. Paris, 2nd series, **30**: 125–133.
- Haddad, C.F.B., Prado, C.P.A. (2005): Reproductive modes in frogs and their unexpected diversity in the Atlantic forest of Brazil. *BioScience*, **55**(3): 207–217.
- IUCN 2008. (2008): IUCN Red List of Threatened Species. <www.iucnredlist.org>. Downloaded on 08 February 2009.
- Jennions, M.D., Backwell, P.R.Y., Passmore, N.I. (1992): Breeding behaviour of the African frog, *Chromantis xerampelina*: multiple spawning and polyandry. *Anim. Behav.*, **44**: 1091–1100.
- Keller, A., Rödel, M. O., Linsenmair, K.E., Grafe, T.U. (2009): The importance of environmental heterogeneity for species diversity and assemblage structure in Bornean stream frogs. *J. Anim. Ecol.*, **78**(2): 305–314.
- Kentwood, D.W. (2007): *The ecology and behavior of amphibians*. The University of Chicago Press, pp. 1148.
- Kok, D., Du Preez, L., Channing, A. (1989): **Channel construction by the African bullfrog: another anuran parental care strategy.** *J. Herpetol.*, **23**: 435–437.
- Lamotte, M., Perret, J.-L. (1961): **Les formes larvaires de quelques espèces de *Leptopelis*: *L. aubryi*, *L. viridis*, *L. anchietae*, *L. ocellata*, et *L. calcaratus*.** Bull. Inst. fond. Afr. noire, Sér. A, **23**: 855–885.
- Lamotte, M., Perret, J.-L. (1963): **Contribution à l'étude des batraciens de l'Ouest africain XIV—le développement direct de l'espèce *Arthroleptis poecilnotus* Peters.** Bull. Inst. fond. Afr. noire, Sér. A, **25**: 277–284.
- Largen, M.J. (1977): The status of the genus *Leptopelis* (Amphibia: Anura: Hyperoliidae) in Ethiopia, including description of two new species. *Monit. Zool. Ital.*, n.s. suppl. **9**: 85–136.
- Lutz, B. (1947): Trends toward non-aquatic and direct development in frogs. *Copeia*, **1947**: 242–252.
- Martin, A. (1967): Australian anuran life histories: Some evolutionary and ecological aspects. In: Weatherly, A. H. (ed), *Australian inland waters and their fauna*, pp. 175–191. Canberra: Australian National University Press.
- Mazibuko, L.C.J., Poynton, J.C. (2004): ***Nothophryne broadleyi***. In: IUCN 2009. IUCN Red List of Threatened Species. Version 2009.1. <www.iucnredlist.org>. Downloaded on 16 June 2009.
- McDiarmid, R.W. (1994): Amphibian diversity and natural history: An overview. In: Heyer et al. (eds.), *Measuring and monitoring biological diversity: Standard methods for amphibians*. Washington (DC): Smithsonian Institution Press.
- McDiarmid, R. W., Altig, R. (1999): *Tadpoles, the biology of anuran larvae*. The University of Chicago Press, Chicago and London.
- Mercurio V. (2009): *Amphibians of Malawi, an analysis of their richness and community diversity in a changing landscape*; Unpublished PhD thesis, Goethe-University, Frankfurt am Main, Germany.
- Passmore, N.I., Carruthers, V.C. (1995): *South African frogs. A complete guide*. Johannesburg, South Africa: University of the Witwatersrand Press.
- Perotti, M.G. (1997): Modos reproductivos y variables reproductivas cuantitativas de un ensamble de anuros del Chaco semi-árido, Salta, Argentina. *Rev. Chil. Hist. Nat.*, **70**: 277–788.
- Pickersgill, M. (1984): Three new *Afraxalus* (Anura: Hyperoliidae) from south-eastern Africa. *Durban Mus. Novit.*, **13**: 203–220.
- Pickersgill, M. (2007): *Frog Search, Results of Expeditions to Southern and Eastern Africa*. Frankfurt Contributions to Natural History 28, Edition Chimaira, Frankfurt.
- Prado, C.P.A., Uetanabaro, M., Haddad, C.F.B. (2005): **Breeding activity patterns, reproductive modes, and habitat use by anurans (Amphibia) in a seasonal environment in the Pantanal, Brazil.** *Amphibia-Reptilia*, **26**(2): 211–221.
- Rödel, M.-O. (1996): *Amphibien der westafrikanischen Savanne*. Edition Chimaira: Frankfurt.
- Rödel, M.-O. (2000): *Herpetofauna of West Africa. Vol. 1. Amphibians of the West African Savanna*. Edition Chimaira: Frankfurt.
- Rödel, M.-O., Spieler, M., Grabow, K., Böckheler, C. (1995): *Hemisus marmoratus* (Peters, 1854) (Anura: Hemisotidae), Fortpflanzung Strategie eines Savannenfrosches. *Bonn. Zool. Beitr.*, **45**: 191–207.
- Salthe, S.N., Duellman, W.E. (1973): Quantitative constraints associated with reproductive mode in anurans. In: Vial, J. L. (ed.), *Evolutionary biology of the anurans*. Columbia: University of Missouri Press.
- Salthe, S.N., Mecham, J.S. (1974): Reproduction and courtship patterns. In: Lofts, B. (ed), *Physiology of the Amphibia*, Vol 2, pp. 309–521. New York: Academic Press.
- Schiotz, A. (1999): *Treefrogs of Africa*. Edition Chimaira, Frankfurt am Main, pp. 350.
- Stevens, R.A. (1972): A new species of *Ptychadena* (Amphibia: Ranidae) from southern Malawi. *Arnoldia (Rhodesia)*, **5**(38): 1–14.
- Stewart, M.M. (1967): *The Amphibians of Malawi*. State University Press, New York.
- Thibaudeau, G., Altig, R. (1999): **Endotrophic anurans. Development and evolution**. In: McDiarmid, R.W. and Altig, R. (eds.), *The biology of anuran larvae*, pp. 170–188.
- Van Dijk, D.E. (1971): Anuran ecology in relation particularly to ovoposition and development out of water. *Zool. Afr.*, **6**: 119–132.
- Van Dijk, D.E. (1972): The behaviour of southern African anuran tadpoles with particular reference to their ecology and related external morphological features. *Zool. Afr.*, **7**: 49–55.
- Van Dijk, D.E. (1977): Habitat and dispersal of southern African anura. *Zool. Afr.*, **12**: 169–181.
- Vences, M., Köhler, J. (2008): Global diversity of amphibians (Amphibia) in freshwater. *Hydrobiologia*, **595**: 569–580.
- Wager, V.A. (1926): Breeding habits and life histories of some Transvaal Amphibia. *Trans. Roy. Soc. South Africa*, **13**: 163–174.
- Wager, V.A. (1960): The blaasop or rainfrog. *Afr. Wild. Life*, **14**: 235–240.
- Wager, V.A. (1965): *The frogs of South Africa*. Capetown, South Africa: Purnell and Sons.
- Wager, V.A. (1986): *Frogs of South Africa, their fascinating life stories*. Craig (Delta), pp. 183.

